

Journal Pre-proofs

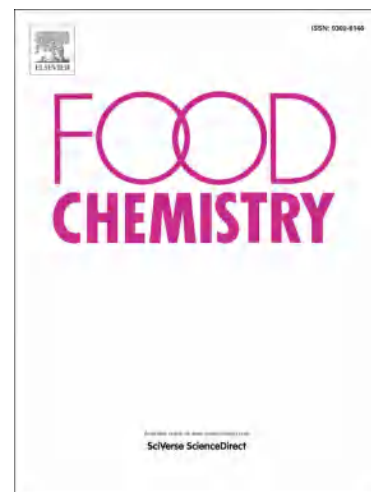
Preparation and FTIR, Raman and SEM characterizations of Konjac Glucomannan-KCl electrogels

Li-Xia Wang, Anng-Ruei Lee, Yi Yuan, Xiu-Mei Wang, Ting-Jang Lu

PII: S0308-8146(20)31151-1
DOI: <https://doi.org/10.1016/j.foodchem.2020.127289>
Reference: FOCH 127289

To appear in: *Food Chemistry*

Received Date: 20 August 2019
Revised Date: 28 May 2020
Accepted Date: 7 June 2020



Please cite this article as: Wang, L-X., Lee, A-R., Yuan, Y., Wang, X-M., Lu, T-J., Preparation and FTIR, Raman and SEM characterizations of Konjac Glucomannan-KCl electrogels, *Food Chemistry* (2020), doi: <https://doi.org/10.1016/j.foodchem.2020.127289>

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2020 Published by Elsevier Ltd.

Preparation and FTIR, Raman and SEM characterizations of Konjac Glucomannan-KCl electrogels

Li-Xia Wang^{a, b, c}, Anng-Ruei Lee^b, Yi Yuan^d, Xiu-Mei Wang^a, Ting-Jang Lu^{b, *}

^a School of Environmental and Biological Engineering, Putian University, Putian 351100, China

^b Graduate Institute of Food Science and Technology, Taiwan University, No.1, Sec.4, Roosevelt Road, Taipei 10617, Taiwan, ROC

^c Fujian Provincial Key Laboratory of Ecology-Toxicological Effects & Control for Emerging Contaminants, Putian University, Putian 351100, China

^d School of Food Science and Engineering, South China University of Technology, Guangzhou 510641, China

ABSTRACT

Konjac glucomannan (KGM) electrogels were successfully prepared under alternating current (AC) in the presence of potassium chloride (KCl). The structure of the gels was studied using Fourier transform infrared spectroscopy (FTIR), Raman spectroscopy, and scanning electron microscopy (SEM). A single-factor experiment was performed to optimize the preparation of the gels. Our results showed that KGM was degraded under AC and

partially deacetylated. KGM and KCl formed the structure $\text{K}^+ \dots \overset{\text{G/M}}{\text{OH}} \dots \text{Cl}^-$, and electrogels with porous structures retained some acetyl groups. Furthermore, as the KCl concentrations, voltages, time, and KGM concentrations increased, the viscoelastic moduli of the gels increased; the moduli decreased when the KCl concentrations, voltages, and time exceeded critical values.

Keywords: Konjac Glucomannan (KGM); Potassium chloride (KCl); Alternating current (AC) electric field; Gel; Structural characterization; Preparation conditions

1. Introduction

Gels are used widely in food products and packaging films, and they are used as additives, matrices, and pharmaceutical and nutrient carriers (Niu, Xia, Li, Wang, & Yu, 2019; Luo, Teng, Wang, & Wang, 2013). The major factor defining the properties of gels is the preparation method, which also determines their applications. Konjac glucomannan (KGM) is a naturally occurring neutral polysaccharide extracted from *Amorphophallus konjac* C. Due to its good film and gel formation capacity, biocompatibility and biodegradability, KGM has been used in the food, medical, environmental, biotechnical and material fields (Zhang, Xie, & Gan, 2005; Zhu, 2018).

For instance, it can form thermally irreversible gels in the presence of an alkaline coagulant (Zhou, Jiang, Perkins, & Cheng, 2018). Alkali facilitates deacetylation and decreases the solvation of KGM chains. KGM transits from random coil configurations to more stiff configurations accompanied by self-assembly to aggregate with one another and form junction zones. The molecular forces responsible for gel formation are thought to be hydrogen bonding and hydrophobic interactions. KGM can also form thermally reversible gels with borate (Gao, Guo, & Nishinari, 2008). The gel network is formed through the crosslinking reaction between borate ions and the *cis*-diol hydroxyl groups on the mannose units of KGM chains. However, boron ions are toxic and non-food grade, which greatly limits the practical applications of the gel in food. KGM could also form reversible complex gels with other polysaccharides or proteins, such as xanthan, carrageenan, and agarose, through intermolecular hydrogen interactions and synergistic effects (Brenner, Tuvikene, Fang, Matsukawa, & Nishinari, 2015). Furthermore, a novel low-alkali alcogel was prepared by mixing ethanol with KGM aqueous solution containing a

38 certain concentration of Na_2CO_3 (Zhou, Wu, Tian, Li, Zhu, Zhao, & Cheng, 2020). The use of alcohol as an
39 antisolvent reduced the solubility of the KGM solute and resulted in the rapid precipitation of the solute or, in
40 some special cases, gelation. Our team previously utilized low-voltage direct current (DC) to induce the formation
41 of KGM electrogels in the presence of tungstate ions. The preparation parameters, rheological properties, and
42 texture properties of these gels were studied, and a mechanism for formation was proposed (Wang, Jiang, Lin,
43 Pang, & Liu, 2016; Wang, Zhuang, Li, Pang, & Liu, 2016).

44 It has been reported that AC can control the orientation of molecules or create crosslinking by
45 dielectrophoresis forces. For instance, Zhuo Chen et al. (2005) demonstrated controllable interconnections of
46 single-walled carbon nanotubes between two electrodes under AC (2.5×10^5 V/m, 5 MHz), and the interconnected
47 carbon nanotubes were parallel with the electric flux. Hermanson et al. (2001) reported that metallic nanoparticles
48 could assemble into microwires in aqueous suspension by dielectrophoresis, and then, the nanoparticles aggregate
49 to extend the wire in the direction of the field gradient with an alternating voltage (50 to 250 V, 50 to 200 Hz)
50 applied to the planar electrodes (resulting in a field intensity of ~ 250 V/cm). However, these studies focused on
51 the assembly or aggregation of small molecules driven by AC with high voltage or high frequency. Whether AC
52 can induce the reorientation, crosslinking, or aggregation of biomacromolecules, such as KGM, to form gels has
53 not been reported. Based on the reports that AC can induce the aggregation of small molecules and our previous
54 research on KGM-tungsten gels induced by DC, we examined the potential of fabricating a new KGM electrogel
55 under AC. Thus, the aim of this study was to utilize an AC electric field to induce KGM to self-assemble into
56 three-dimensional gel networks and examine the structural and rheological characteristics of the gels.

57 58 **2. Materials and methods**

59 *2.1. Materials*

60 KGM powder was purchased from Sanai Konjac Biotechnology Co., Ltd. (Zhaotong, China). KCl was
61 analytical grade and purchased from Cuilin Huabo Instrument Co., Ltd. (Zhangzhou, China).

62 *2.2. Preparation of KGM-KCl electrogel under AC electric field*

63 KGM-KCl electrogels were prepared according to Wang et al. (2016) with some modifications. Briefly, 0.5 ~
64 0.9% KCl was added to KGM hydrosol (0.5~1.1%). Carbon rods were inserted into the mixture, and an AC
65 (21~25 V) was applied for at least 35 min. The distance between the two rods was fixed at 4 cm. Optimal gel
66 formation was achieved using a mono-factor experiment, using elastic modulus as the evaluation parameter.

67 *2.3. Fourier transform infrared (FTIR) spectroscopy*

68 FTIR spectra of KGM powder, lyophilized KGM hydrosol treated by an AC electric field for a certain time
69 (KGM-AC) and frozen-dried gel samples were recorded on a Nicolet iS10 FTIR spectrometer (Thermo Fisher
70 Scientific Inc., WI, USA) in the range of 400 - 4000 cm^{-1} .

71 *2.4. Raman spectroscopy*

72 Raman spectra were recorded with a Labram Aramis confocal microprobe analyser (Horiba Jobin-Yvon,
73 Longjumeau, France) equipped with an Olympus microscope and a CCD cooled according to Wang, Jiang, Lin,
74 Pang and Liu (2016).

75 *2.5. Scanning electron microscopy (SEM)*

76 KGM powder, lyophilized KGM hydrosol, lyophilized KGM hydrosol treated by an AC electric field for a
77 certain time (KGM-AC) and lyophilized KGM-KCl gels were attached to the aluminium plate through
78 double-sided conductive carbon tabs. After being coated with gold in an SCD-500 ion sputter (Hitachi Ltd.,
79 Hitachi, Tokyo, Japan), the samples were observed by a Quanta250/Quanta430 Thermal Field Emission Scanning
80 Electron Microscope (FEI Co., Hillsboro, OR, USA).

81 2.6. Rheological measurements

82 Steady flow measurements and dynamic oscillatory tests were performed on an AR-2000 ex stress-controlled
83 rheometer (TA Instrument, New Castle, DE, USA) equipped with a Peltier device for temperature control using a
84 plate-plate geometry (diameter 40 mm for sol samples, 20 mm for gel samples, gap 0.2 mm). KGM and
85 KGM-KCl sols were loaded on the plate, allowed to equilibrate at 25 °C for 2 min, and then subjected to
86 increasing shear rates from 1 to 200 s⁻¹ (Wang, Xiong, & Sato, 2017). The apparent viscosity (η_a) was recorded as
87 a function of shear rate. The fresh gel samples were also equilibrated at 25 °C for 2 min before rheological
88 measurements. The linear viscoelastic region (LVR) for gel samples was determined by performing amplitude
89 sweep measurements (0.01-5%) at constant frequency (1 Hz) and 25 °C. Frequency sweep tests at a constant strain
90 of 0.5% were carried out to determine the viscoelastic nature of the KGM-KCl electrogel. Under this strain limit,
91 the gel network structure will not be destroyed by the measurements. All measurements were performed at least in
92 triplicate.

93 3. Results and discussion

94 3.1. Preparation of KGM-KCl electrogel under AC electric field

95 When an AC electric field was applied on a certain volume of 0.5% KGM hydrosol, no gel formation was
96 observed, even with an electric treatment of 45 min. There was also no heat release phenomenon or gas
97 production during the course of electric treatment. No gel formed when KGM hydrosol contained sodium
98 tungstate treated by AC. However, white gels were produced at the gas/liquid interface when AC was utilized to
99 process 0.5% KGM hydrosol containing a certain amount of KCl. Moreover, bubbles were produced, and heat was
100 released during the treatment. The bubbles were ascribed to the production of H⁺ and OH⁻ from water electrolysis
101 during the course of electric treatment, resulting in the release of hydrogen and oxygen gases. It also occurred in
102 the process of water electrolysis caused by a DC (Wang, Zhuang, Li, Pang, & Liu, 2016). Heat release was due to
103 the violent movement of ions in the AC electric field. It should be noted that when the KCl concentration was
104 above 0.9%, no gel formed. At this time, the KGM hydrosol began to boil, and its fluidity increased under high
105 temperature caused by ion movements. In addition, it can be observed that the KGM sol became thinner, as KGM
106 could not withstand temperatures above 80 °C (Wang, Jiang, Lin, Pang, & Liu, 2016). Therefore, a suitable
107 concentration of KCl was necessary for the formation of KGM electrogels under an AC electric field. On the other
108 hand, when the voltage applied was below 21 V or above 26 V, there was also no gel formation, indicating that the
109 application of a suitable voltage was another important factor that determined the gel formation. Overall, an
110 appropriate KCl concentration and voltage are essential conditions for the formation of KGM electrogels.

111 3.2 FTIR spectroscopy

112 FTIR spectra of native KGM powder, KGM-AC and KGM-KCl gel are shown in Fig. 1. The strong
113 absorption at 3346.99 cm⁻¹ corresponded to the stretching vibration of -OH. The peak at 2884.14 cm⁻¹

114 corresponded to the C-H stretching vibrations in the $-\text{CH}_2$ or $-\text{CH}_3$ groups. The stretching vibration of the carbonyl
 115 at 1724.12 cm^{-1} corresponded to characteristic acetyl groups of KGM molecules (Li, Ma, Chen, He, & Huang,
 116 2018). The absorption band at 1640.23 cm^{-1} corresponded to the C-O stretching vibrations related to $-\text{OH}$ (Ye, Jin,
 117 Huang, Hu, Li, Li, & Li, 2017). The absorption peak at 1022.13 cm^{-1} corresponded to C-O-C vibrations (Li, Ji, Li,
 118 Zhang, Xiong, & Sun, 2017). The absorption peaks at 898.70 cm^{-1} and 805.65 cm^{-1} corresponded to the stretching
 119 vibrations of mannose units (Wang, Wu, Xiao, Kuang, Corke, Ni, & Jiang, 2017).

120 As shown in Fig. 1, some differences were found among the spectra. The chemical structures of KGM-AC
 121 and KGM-KCl electrogel were similar to that of native KGM. When compared with the spectrum of native KGM,
 122 the peak at 3358.42 cm^{-1} in KGM-AC became distinctly weaker and wider, indicating that the $-\text{OH}$ vibrations
 123 were affected. In addition, the peak in native KGM at 3346.99 cm^{-1} shifted to a higher frequency of 3358.42 cm^{-1}
 124 in KGM-AC, which could be attributed to the complex vibrational stretches associated with free, inter- and
 125 intramolecular hydroxyl groups (Wang, Zhou, Wang, & Li, 2015). The higher the peak frequency was, the weaker
 126 the interaction was (Sun, Li, Dai, Ji, & Xiong, 2014). Li et al. also reported that the characteristic bands at
 127 $3500\text{--}3300\text{ cm}^{-1}$ of short linear glucan (SLG) nanoparticles shifted to a lower wavelength in SLG/soy protein
 128 isolate (SPI) nanoparticles, indicating that the interaction force of intermolecular hydrogen bonds between SLG
 129 and SPI molecules was enhanced (Li, Ji, Li, Zhang, Xiong, & Sun, 2017). These results all showed that the
 130 vibration of $-\text{OH}$ in KGM-AC was affected. Meanwhile, the peak at 1022.13 cm^{-1} in native KGM shifted to a
 131 higher frequency of 1025.46 cm^{-1} in KGM-AC and became weaker, suggesting the breaking of glycosidic bonds
 132 (Wang, Zhou, Wang, & Li, 2015). From the above analysis, we could conclude that KGM degraded under an AC
 133 electric field. In addition, the stretching peak of the carbonyl at 1724.12 cm^{-1} in KGM-AC decreased, indicating
 134 that the characteristic acetyl groups of KGM molecules decreased; this decrease might have been due to a certain
 135 degree of deacetylation by hydroxide ions, which derived from water electrolysis.

136 However, the peaks of $-\text{OH}$ and C-O-C stretching vibrations in the KGM-KCl gel strengthened again. It is
 137 plausible that electrostatic rejection and electrostatic attraction resulting from K^+ and Cl^- on hydroxyl groups
 138 might be responsible for the strengthening of the vibration. The interactions between hydroxyl and ions included
 139 oxygen atoms of hydroxyl groups interacting with metal ions and hydrogen atoms having a weak interaction with
 140 anions to form the structure of $\text{M}^{\text{n}+} \dots \text{OH} \dots \text{anion}^{\text{n}-}$. The interactions between the hydroxyl groups of citrus pectin
 141 and monovalent cations, such as Na^+ and K^+ , were also elucidated by Wang et al. (Wang, Wan, Chen, Guo, Liu, &
 142 Pan, 2019). In our system, hydroxyl groups of KGM molecules interacted with K^+ and Cl^- to form the structure of

143 $\begin{array}{c} \text{G/M} \\ | \\ \text{K}^+ \dots \text{OH} \dots \text{Cl}^- \end{array}$. G represents glucose, and M represents mannose. Thus, the hydrogen bond interactions in the gel
 144 were decreased. Combined with the fact that the gel formed at the gas/liquid interface, we believed that the main
 145 forces of gel formation were electrostatic and hydrophobic interactions instead of hydrogen bonds.

146 The stretching peak of the carbonyl at 1724.12 cm^{-1} indicated that the characteristic acetyl groups of KGM
 147 molecules remained in the KGM-AC and KGM-KCl electrogels. No new characteristic peaks appeared in
 148 KGM-AC and KGM-KCl gels, indicating that AC electric field treatment did not change the main structure of
 149 KGM molecules or produce new functional groups.

150 3.3 Raman analysis

151 Fig. 2 shows Raman spectra for KGM powder, KGM-AC and KGM-KCl electrogel in the wavenumber
152 range of 0-3500 cm^{-1} . The weak band at 3360.18 cm^{-1} corresponded to the O-H stretching vibration (Mu et al.,
153 2018). The strong band at 2888.84 cm^{-1} corresponded to the C-H stretching modes (Zhang, Yang, Xu et al., 2018).
154 The weak peak at 1657.72 cm^{-1} corresponded to the vibration of carbonyl (C=O), and the peak at 1453.71 cm^{-1}
155 corresponded to the vibration of methyl (-CH₃) in the acetyl group (Jian, Siu, & Wu, 2015). The bands at 1117.38
156 cm^{-1} and 1085.27 cm^{-1} corresponded to the C-O-C vibration from the glycosidic bonds of KGM molecules (Mu et
157 al., 2018).

158 In comparison with the Raman spectra of KGM and KGM-AC, almost all the typical characteristic peaks
159 were weakened. The peaks at 1124 cm^{-1} and 1084.32 cm^{-1} decreased, indicating that the vibration of C-O-C in
160 KGM-AC was affected. That is, the glycosidic bonds broke. In addition, -OH, C-H and C=O vibrations also
161 decreased in the spectrum of KGM-AC, suggesting that KGM degraded under an AC electric field and partially
162 deacetylated due to the increase in local hydroxide ion concentration.

163 The O-H vibrational band at 3360.18 cm^{-1} recovered in the spectrum of the KGM-KCl gel, which could be
164 explained as the result of electrostatic interactions induced by the adsorption of K⁺ and Cl⁻ ions on hydroxyl
165 groups of KGM (Wang, Shen, Liu et al., 2018). This explanation is supported by Qiang Sun and his collaborator.
166 They investigated the interactions among monovalent and divalent cations, chloranion and water in NaCl-H₂O,
167 KCl-H₂O, CaCl₂-H₂O, MgCl₂-H₂O and NaCl-KCl-CaCl₂-H₂O systems by Raman spectra and found that anions
168 interacted with water molecules on the hydrogen donor side of the molecule, whereas cations interacted through
169 lone pairs (acceptor side) (Sun, Zhao, Li, & Liu, 2010). As a result, hydrogen bond interactions decreased
170 correspondingly.

171 Furthermore, Raman bands near 1450 cm^{-1} and 2800-3000 cm^{-1} corresponded to C-H bending and stretching
172 vibrations, respectively (Herrero et al., 2008). As shown in Fig. 2, the C-H bending and stretching vibrations of
173 KGM were located at 1453.71 cm^{-1} and 2888.84 cm^{-1} , respectively, while KGM-KCl gel had stronger vibration
174 intensities near 1381.74 cm^{-1} and 2899.8 cm^{-1} . This might be because methyl- or methylene groups of the
175 KGM-KCl gel were exposed to the polar environment of the KCl solution. The changes in location, intensity and
176 band area, which are assigned to the vibrations of the C-H bond, indicate changes in the environment of C-H
177 groups, which may be related to hydrophobic interactions (Xu, Han, Fei, & Zhou, 2011). Larsson & Rand also
178 reported that the intensity of the 2930 cm^{-1} band increased with increasing polarity of the environment around
179 hydrocarbon chains (Larsson & Rand, 1973). In addition, the higher intensity of C-H stretching vibrations at
180 2899.8 cm^{-1} implied that the -CH hydrophobic groups of KGM in the gel were not exposed, which could be
181 ascribed to the more compact gel structure (Xue, Qian, Kim, Xu, & Zhou, 2018). According to the above analysis,
182 combined with experimental phenomena, we thought the main forces for gel formation and gel stability were
183 electrostatic and hydrophobic interactions. These results all coincided with the FTIR analysis.

184 3.4 SEM observations

185 Fig. 3 shows SEM images of native KGM powder (a), lyophilized KGM hydrosol (b), KGM-AC (c-f) and
186 lyophilized KGM-KCl gels (g-i) with different magnifications. KGM powder presented a regular microfibre shape
187 with many short chain branches between the tightly arranged backbone chains. The microfibrils were glued to
188 each other, which might be due to intra- and intermolecular hydrogen bond formation between KGM molecules.

189 The bonds played an important role in stabilizing the microfibre chain structure of KGM (Fig. 3a).

190 The morphology of the freeze-dried KGM hydrosol is shown in Fig. 3b. KGM presented a disorganized
191 flocculation instead of the original regular microfibre structure after freeze-drying. This was because water
192 molecules bound and filled between KGM molecular chains by many intermolecular hydrogen bonds evaporated
193 during vacuum freeze drying, leaving the swollen KGM molecular chain frozen and fixed in its original position
194 to form a scaffold structure. In addition, some KGM micelles were thicker and some were thinner due to the
195 strong interaction between KGM molecules, which was caused by hydrogen bonds.

196 Distinct changes were found in the morphology of KGM-AC compared with that of KGM powder and
197 lyophilized KGM hydrosol. As shown in Fig. 3 (c-f), the regular microfibrils of KGM and its scaffold structure
198 disappeared. Instead, KGM presents a flaky and uneven layered structure. This was because KGM microfibrils
199 were fractured under AC electric field treatment, and short KGM molecular chains could not wrap water
200 molecules to form scaffold structures. The results were in accordance with the analysis of FTIR and Raman
201 spectra. In addition, it can be seen from the image with high magnification (Fig. 3f) that there are obvious
202 breaking of KGM molecular chains. We also found that the lyophilized KGM hydrosol treated by an AC electric
203 field was fragmented into several pieces with different sizes rather than a whole piece of sponge-like product by
204 experimental observations.

205 Fig. 3 (g-i) shows the morphological structure and surface images of AC electric field-induced KGM-KCl
206 gels. It can be seen from the SEM photographs that KGM-KCl electrogels exhibited a porous structure, allowing
207 the gel to maintain certain elastic characteristics. It possessed a rough and irregular surface with many folds. The
208 walls of the gels were coarse, and the pore size distribution was not homogeneous.

209 3.5. Steady shear measurements of KGM sols and KGM-KCl mixed sols

210 To explore the effect of the AC electric field on the molecular chains of KGM, steady shear measurements
211 were conducted. Apparent viscosities of KGM sols and KGM-KCl complex hydrosols without and with electric
212 treatment were recorded on an advanced AR-2000 ex rheometer. The mixed sol was treated for 25 min by
213 applying an AC electric field to not form a gel. In addition, according to our knowledge, the formation of
214 electrogel is a process of aggregation of KGM molecules between electrodes; thus, the homogeneous KGM-KCl
215 sol becomes a nonuniform system after electroprocessing for a certain time. Therefore, KGM composite sols for
216 determination were sampled from different points, including from the centre of the beaker, from the vicinity of the
217 beaker wall and from the vicinity of the electrode. The flow curves are presented in Fig. 4. From Fig. 4, we can
218 see that the apparent viscosities of all the samples, including KGM sol and KGM-KCl mixtures with and without
219 electric treatment, all decreased as the shear rate increased, demonstrating that the sols exhibited non-Newtonian
220 fluid properties (Ma, Zhu, & Wang, 2019). Furthermore, the apparent viscosities of the KGM sol (Fig. 4a) and
221 KGM-KCl mixture (Fig. 4b) treated by an AC electric field were all lower than that of the sol without electric
222 treatment, indicating the breaking of KGM molecular chains after electric treatment in water or KCl solution
223 (Tatirat, Charoenrein, & Kerr, 2012). That is, AC electric field treatment was found to be able to reduce KGM
224 solution viscosity in a simple and controllable manner, and it could be an effective degradation method for KGM
225 and other polysaccharides. This result was in good agreement with the findings from the FTIR, Raman and SEM
226 analyses.

227 3.6. Frequency sweep

228 3.6.1. Effect of KCl concentration on G' and G'' of KGM-KCl gel

229 To explore the optimum conditions for preparing KGM-KCl electrogel, the effects of four factors, including
230 KCl concentration, electric treatment time, voltage and KGM concentration, on the rheological properties of the
231 gels were studied. First, considering the importance of KCl in gel formation, we studied the effect of KCl
232 concentration on the rheological properties of the gels. To examine the frequency dependence of G' and G'' of the
233 KGM-KCl electrogel, frequency sweep tests were carried out using small amplitude oscillatory shear. As shown
234 in Fig. 5a, in the high frequency area, G' and G'' of the gels increase only slightly with frequency, suggesting the
235 formation of a good gel network structure in the samples. This slight increase also indicated their abilities to resist
236 external deformation (Wang, Jiang, Lin, Pang, & Liu, 2016). In addition, the elastic modulus (G') of all the
237 samples prepared with different concentrations of KCl dominates the viscous modulus (G''), which indicates the
238 gel-like nature of the products (Li, Ye, Zhou, Lei, & Zhao, 2019). By comparing the changes in moduli of the gels
239 prepared with different concentrations of KCl (c_k), we found that when c_k increased from 0.5% to 0.7%, G' and G''
240 increased. This might be because the adsorption of chloride ions on the -OH groups of KGM molecules depends
241 on chloride ion concentrations. As the concentration of Cl^- ions increases, more adsorption occurs (Bagheri,
242 Nazari, Sanjayan, & Duan, 2018). KGM was more charged with more KCl due to the adsorption of K^+ and Cl^-
243 ions, and thus, the charged KGM molecular chains interpenetrated each other to form network structures under an
244 AC electric field. When c_k was 0.7%, the KGM sol started to boil in the last few minutes of AC electric field
245 processing. Under this circumstance, the gel could still form. The moderate boil not only caused the gel to form at
246 the gas/liquid interface but could also remove some of the gases from the gel. Therefore, the moduli of the gels
247 were relatively higher. However, G' and G'' decreased when c_k exceeded 0.7%. This was because the greater the
248 amount of KCl added, the greater the heat release during the application of the AC electric field. As a result, on
249 the one hand, KGM molecular chains broke severely under high temperature. On the other hand, the high heat and
250 temperature destroyed the stability of the formed gel. When c_k was above 0.9%, it was no longer possible to
251 produce gels. This was because the added KCl was too much, and KGM sol boiled too violently to form the gel.
252 The whole gelation process reflected the vital role of KCl and its suitable concentration in gel formation.
253 Therefore, we chose a concentration of 0.7% KCl.

254 3.6.2. Effect of electric processing time on G' and G'' of KGM-KCl gel

255 Second, to explore the importance of the application of an AC electric field on gel formation, we studied the
256 influence of electric processing time on the rheological properties of KGM-KCl gels. The results are shown in Fig.
257 5b. When an AC electric field was applied for 35 min, the gel formed. Notably, no gel formed when the electric
258 treatment time was less than 35 min. It can be seen from Fig. 5b that when the electric treatment time is increased
259 from 40 min to 50 min, G' and G'' correspondingly increase with G' being larger than G'' . This is probably because
260 when the voltage, KCl concentration and KGM concentration are fixed, the time needed for the intercutting of
261 KGM molecular chains extends with the increase of electric treatment time, resulting in the formation of a more
262 compact gel network. When the electric treatment time was increased from 50 min to 60 min, G' and G''
263 correspondingly decreased, indicating a decrease in gel strength. When the electric treatment time exceeded 60
264 min, the formed gel dissolved. This might be because as the electric processing time extended, the stability of the

265 gel decreased under the high temperature caused by the heat release, which was generated by the movement of
266 ions. From the above, KGM-KCl electrogel can form in at least 35 min of electric treatment. This differs from our
267 previous reports about the preparation of KGM-tungsten electrogels, which can form in 3 min of DC electric
268 processing (Wang, Jiang, Lin, Pang, & Liu, 2016; Wang, Zhuang, Li, Pang, & Liu, 2016). These experiments
269 highlighted the importance of the application of an AC electric field in KGM-KCl electrogel formation. Therefore,
270 an electric treatment time of 50 min was chosen in the following experiments.

271 3.6.3. Effect of voltage on storage modulus (G') and loss modulus (G'') of KGM-KCl gel

272 The effect of voltage on the rheological properties of KGM-KCl gels is shown in Fig. 5c. As shown in the
273 figure, voltage significantly affected the rheological properties of the gels. As the voltage increased by 1 V, the G'
274 and G'' of the gels changed greatly, and the effect of the voltage was similar to the effects of KCl concentration
275 and electric treatment time. In particular, when the voltage increased from 21 V to 23 V, G' and G'' of the gel
276 increased, with G' being larger than G'' . This might be because with increasing voltage, the electric field strength
277 increased and the movement of the charged KGM molecular chains accelerated, resulting in more intercutting of
278 KGM chains under a fixed electric processing time; thus, the strength of the formed gel increased. However, when
279 the voltage increased from 23 V to 25 V, G' and G'' of the gel decreased. This might be because the motion of
280 some unadsorbed K^+ and Cl^- ions strengthened under high electric field strength, resulting in substantial heat
281 release, which reduced the stability of the formed gel. On the other hand, KGM molecular chains broke more
282 severely under high temperatures, as we mentioned above, and under high electric field strength, which has been
283 verified by SEM observations and steady shear measurements. When the voltage was below 21 V or exceeded 25
284 V, no gel appeared. This might be because either the electric current was too low to drive the motion and
285 entanglement of charged KGM molecular chains or because the electric current and heat released were too high to
286 form the gel. Additionally, a higher voltage could lead to the KGM sol boiling violently. The effect of the voltage
287 on the moduli of the gels and the gel formation suggested that the application of an appropriate voltage was
288 critical to the formation and stability of a gel. This finding also indicated that AC electric treatment was one of the
289 necessary conditions for the formation of the electrogel. Therefore, we chose 23 V as the voltage in the following
290 experiments.

291 3.6.4. Effect of KGM concentration on G' and G'' of KGM-KCl gel

292 Finally, the effect of KGM concentrations (c_{KGM}) on G' and G'' of KGM-KCl electrogels was studied. The
293 results are presented in Fig. 5d. It can be seen from Fig. 5d that G' and G'' of the KGM-KCl gel increase with
294 increasing c_{KGM} from 0.5% to 1.1%. This might be because with increasing c_{KGM} , the number of charged KGM
295 molecules involved in interpenetrating and tangling with each other increases. When c_{KGM} reached 1.3%, the
296 KGM sol became thicker, and the mobility of KGM molecular chains was affected. Therefore, we did not prepare
297 the gel at a 1.3% concentration of KGM. The trend of the dependence of G' and G'' on KGM concentration was
298 opposite to that of KGM-tungsten gels we reported earlier (Wang, Jiang, Lin, Pang, & Liu, 2016) because the
299 gelation mechanism of KGM-KCl and KGM-tungsten electrogels was different. For KGM-KCl gels, the charged
300 KGM molecules due to the adsorption of K^+ and Cl^- ions on -OH groups of KGM molecules are the key to
301 forming the gel, while for KGM-tungsten gels, isopoly-tungstic acid ions generated from the transformation of
302 WO_4^{2-} ions cross-link with -OH groups at the C-6 position on KGM under a DC electric field to form a

303 three-dimensional network. Thus, the ratio of isopolytungstic acid ions to KGM is critical to the formation and
304 stability of the gel. Therefore, a concentration of 1.1% KGM was chosen for preparing KGM-KCl gel.

305 It has been reported that gels could form at least 0.5% c_{KGM} in NaOH solution (Luo, He, & Lin, 2013), and
306 for KGM/borax gel, the lowest concentration of KGM studied was also 0.5% (Gao, Guo, & Nishinari, 2008). In
307 our previous publications, gels could form at only 0.3% c_{KGM} in the presence of sodium tungstate under a DC
308 electric field (Wang, Jiang, Lin, Pang, & Liu, 2016; Wang, Zhuang, Li, Pang, & Liu, 2016). In this research, we
309 obtained gels at 0.5% c_{KGM} in the presence of KCl under an AC electric field. This indicates that the application of
310 an AC electric field can also promote gel formation at almost the same low KGM concentrations.

311 Two Chinese patents, namely, CN101328222A and CN101328224A, have been approved for the preparation
312 of *licorice and choerospondiatis* polysaccharide-metal complexes. The metal compounds include Na_2WO_3 ,
313 Na_2SeO_4 , FeCl_2 , CaCl_2 , and KCl, among others. The FTIR spectra of the complexes were also analysed. The
314 results showed that the compounds had the same spectral characteristics as the native polysaccharides. Their
315 characteristic peaks were at $3423\text{-}3419\text{ cm}^{-1}$, $2926\text{-}2847\text{ cm}^{-1}$, $1607\text{-}1621\text{ cm}^{-1}$, $1159\text{-}1132\text{ cm}^{-1}$ and $1098\text{-}1015$
316 cm^{-1} , which corresponded to the vibrations of -OH, -CH₂, -C=O, -C-O-C and -C-O-H, respectively. They also had
317 the characteristic absorption peaks of β - and α - pyranose. However, the preparation method was more
318 time-consuming, complicated, and labourious. It included filtering, dialysis, condensation, alcohol precipitation,
319 sedimentation, washing, drying, etc. Finally, pure polysaccharide-metal materials were obtained.

320 In contrast, gel formation by applying an AC electric field is a much simpler, faster and more effective,
321 controllable and adjustable method. It just requires an electrical current, electric field strength (i.e., voltage and
322 the distance between the electrodes), KCl and KGM concentrations. Gel formation does not require heating. Gels
323 can be obtained at a low concentration of 0.5% KGM with the aid of KCl without adding alkali and other
324 substances. In addition, the gels form under high temperature circumstances, which endows the gels with high
325 thermal stabilities. Compared with the method of DC induction, the use of AC can overcome serious electrolytic
326 hydrolysis and electrochemical corrosion. Gel formation induced by AC requires only nontoxic KCl, without the
327 help of toxic sodium tungstate. The effect of voltage on the formation of KGM-KCl gel induced by AC is greater
328 than that of KGM-tungsten induced by DC (Wang, Jiang, Lin, Pang, & Liu, 2016; Wang, Zhuang, Li, Pang, & Liu,
329 2016). A small change in voltage can affect the formation of KGM-KCl gel and its moduli. However, it takes a
330 longer time for the KGM-KCl electrogel to form under the AC electric field because the formation mechanisms of
331 the two kinds of gels are different. In addition, AC can cause the KGM molecular chains to break, so the gel
332 formed is relatively small and thin, while DC does not cause the KGM molecular chains to break. In the future,
333 the adsorption properties, dielectric properties, electrochemical properties, mechanical properties and other
334 properties of these two kinds of gels should be studied and compared.

335 3.7 The formation mechanism of the KGM-KCl electrogel

336 Based on the above experiments and analysis, we summarized that the formation mechanism of the
337 KGM-KCl electrogel was as follows: K^+ and Cl^- ions adsorb on the hydroxyl groups of KGM molecular chains,
338 and the charged KGM molecular chains run through each other to form a topological entanglement structure
339 between the two electrodes under an AC electric field. Meanwhile, the number of interlocking molecules per unit
340 volume increases due to the breakage of KGM molecular chains under AC. In addition, the AC electric field

341 promotes the ordered orientation of the KGM molecular chain parallel to the direction of the electric field.
 342 Electrostatic and hydrophobic interactions are the main driving forces for gel formation, rather than hydrogen
 343 bonds. A schematic representation of the formation mechanism of the gel is shown in Fig. 6.

344 Conclusion

345 In this study, KGM-KCl electrogels were successfully prepared at 0.5% KGM in the presence of KCl under an
 346 AC electric field. The FTIR, Raman and SEM characterization and rheological properties of the gels were studied,
 347 and the formation mechanism was summarized. The results showed that the AC electric field did not change the
 348 chemical main chain structure of KGM or produce new functional groups. KGM partially deacetylated under an
 349 AC electric field due to the hydroxide ions resulting from electrolytic hydrolysis. In addition, KGM molecular
 350 chains broke under an AC electric field and high temperature. The gels retained some characteristic acetyl groups
 351 of KGM molecules and possessed an ununiform porosity structure. The storage and loss moduli of KGM-KCl gels
 352 were found to increase when the KCl concentration, KGM concentration, voltage and electric processing time
 353 increased but decrease when the KCl concentration, voltage and electric processing time exceeded critical points.

354 KGM and KCl interacted with each other to form the structure of $\begin{matrix} \text{G/M} \\ | \\ \text{K}^+ \cdots \text{OH} \cdots \text{Cl}^- \end{matrix}$. Thus, the charged
 355 KGM inter-penetrated or tangled with each other to form the gel network with the application of an AC electric
 356 field. This novel electrogel will have potential applications in food, material engineering, chemical industries, etc.

357 Acknowledgments

358 This study was supported by the Doctor Scientific Research Foundation of Putian University (2019022),
 359 National Natural Science Foundation of China (31801462), Research and Innovation Projects of Putian University
 360 (2018ZP03), the Education and Research Project of Young and Middle-aged Teachers of Fujian Province
 361 (JT180467), and the Cultivation Program for the Outstanding Young Scientific Research Talents of Fujian
 362 Province (2018).

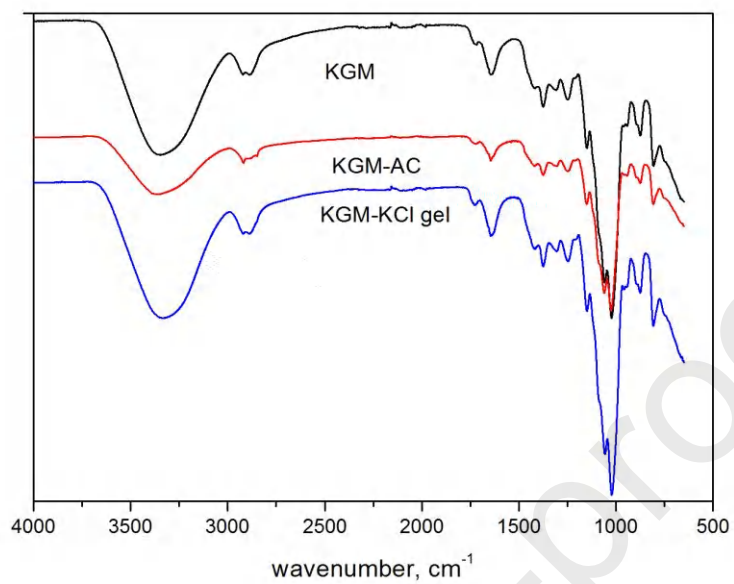
363 References

- 364 Bagheri, A., Nazari, A., Sanjayan, J. G., & Duan, W. H. (2018). Molecular simulation of water and chloride ion
 365 diffusion in nanopores of alkali-activated aluminosilicate structures. *Ceramics International*, *44*, 20723-
 366 20731.
- 367 Brenner, T., Tuvikene, R., Fang, Y. P., Matsukawa, S., & Nishinari, K. (2015). Rheology of highly elastic
 368 iota-carrageenan/ kappa-carrageenan/ xanthan/ konjac glucomannan gels. *Food Hydrocolloids*, *44*, 136-144.
- 369 Chen, Z., Yang, Y. L., Chen, F., Qing, Q., Wu, Z. Y., & Liu, Z. F. (2005). Controllable Interconnection of
 370 Single-Walled Carbon Nanotubes under AC Electric Field. *The Journal of Physical Chemistry. B*, *109* (23),
 371 11420-11423.
- 372 Gao, S. J., Guo, J. M., & Nishinari, K. (2008). Thermoreversible konjac glucomannan gel crosslinked by borax.
 373 *Carbohydrate Polymers*, *72*, 315-325.
- 374 Herrero, A., Carmona, P., Cofrades, S., & Jiménez-Colmenero, F. (2008). Raman spectroscopic determination of
 375 structural changes in meat batters upon soy protein addition and heat treatment. *Food Research International*,
 376 *41*(7), 765-772.
- 377 Hermanson, K. D., Lumsdon, S. O., Williams, J. P., Kaler, E. W., & Velev, O. D. (2001). Dielectrophoretic

- 378 Assembly of Electricly Functional Microwires from Nanoparticle Suspensions. *Science*, 294, 1082-1086.
- 379 Jian, W. J., Siu, K. C., & Wu, J. Y. (2015). Effects of pH and temperature on colloidal properties and molecular
380 characteristics of Konjac glucomannan. *Carbohydrate Polymers*, 134, 285-292.
- 381 Larsson, K., & Rand, R. P. (1973). Detection of changes in the environment of hydrocarbon chains by Raman
382 spectroscopy and its application to lipid-protein systems. *Biochimica et Biophysica Acta (BBA) - Lipids and
383 Lipid Metabolism*, 326(2), 245-255.
- 384 Li, M. Y., Feng, G. P., Wang, H., Yang, R. L., Xu, Z. L., & Sun Y. M. (2017). Deacetylated konjac glucomannan is
385 less effective in reducing dietary-induced hyperlipidemia and hepatic steatosis in C57BL/6 mice. *Journal of
386 agricultural and food chemistry*, 65(8), 1556-1565.
- 387 Li, X. J., Ji, N., Li, M., Zhang, S. L., Xiong, L., & Sun, Q. J. (2017). Morphology and structural properties of
388 novel short linear glucan / protein hybrid nanoparticles and their influence on the rheological properties of
389 starch gel. *Journal of Agricultural and Food Chemistry*, 65 (36), 7955-7965.
- 390 Li, J. W., Ma, J. W., Chen, S. J., He, J. M., & Huang, Y. D. (2018). Characterization of calcium alginate/
391 deacetylated konjac glucomannan blend films prepared by Ca²⁺ crosslinking and deacetylation. *Food
392 Hydrocolloids*, 82, 363-369.
- 393 Li, S. H., Ye, F. Y., Zhou, Y., Lei, L., & Zhao, G. H. (2019). Rheological and textural insights into the blending of
394 sweet potato and cassava starches: in hot and cooled pastes as well as in fresh and dried gels. *Food
395 Hydrocolloids*, 89, 901-911.
- 396 Liu, W. Y., Wang, C. H., Ding, H. T., Shao, J. Y., & Ding, Y. C. (2016). AC electric field induced
397 dielectrophoretic assembly behavior of gold nanoparticles in a wide frequency range. *Applied Surface
398 Science*, 370, 184-192.
- 399 Luo, X. G., He, P., & Lin, X. Y. (2013). The mechanism of sodium hydroxide solution promoting the gelation of
400 konjac glucomannan (KGM). *Food Hydrocolloids*, 30, 92-99.
- 401 Luo, Y. C., Teng, Z., Wang, X. G., & Wang, Q. (2013). Development of carboxymethyl chitosan hydrogel beads
402 in alcohol-aqueous binary solvent for nutrient delivery applications. *Food Hydrocolloids*, 31, 332-339.
- 403 Ma, S. P., Zhu, P. L., & Wang, M. C. (2019). Effects of konjac glucomannan on pasting and rheological properties
404 of corn starch. *Food Hydrocolloids*, 89, 234-240.
- 405 Mu, R. J., Wang, L., Du, Y., Yuan, Y., Ni, Y. S., Wu, C. H., & Pang, J. (2018). Synthesis of konjac glucomannan-
406 silica hybrid materials with honeycomb structure and its application as activated carbon support for Cu(II)
407 adsorption. *Materials Letters*, 226, 75-78.
- 408 Niu, Y. G., Xia, Q., Li, N., Wang, Z. Y., & Yu, L. L. (2019). Gelling and bile acid binding properties of gelatin-
409 alginate gels with interpenetrating polymer networks by double cross-linking. *Food Chemistry*, 270, 223-228.
- 410 Parikka, K., Nikkila, I., Pitkanen, L., Ghafar, A., Sontag-Strohm, T., & Tenkanen, M. (2017). Laccase/TEMPO
411 oxidation in the production of mechanically strong arabinoxylan and glucomannan aerogels. *Carbohydrate
412 Polymers*, 175, 377-386.
- 413 Sam, M., Moghimian, N., & Bhiladvala, R. B. (2016). Field-directed chaining of nanowires: towards transparent
414 electrodes. *Materials Letters*, 163, 205-208.
- 415 Sun, Q., Li, G., Dai, L., Ji, N., & Xiong, L. (2014). Green preparation and characterisation of waxy maize starch

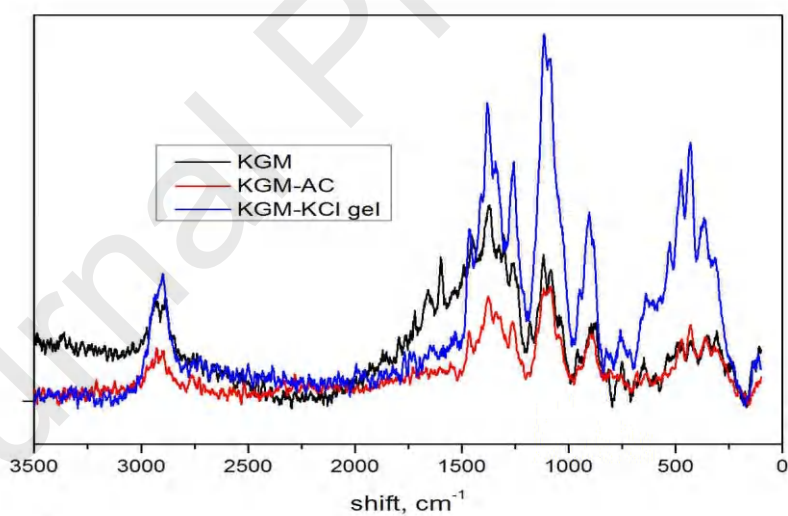
- nanoparticles through enzymolysis and recrystallisation. *Food Chemistry*, 162, 223-228.
- Sun, Q., Zhao, L., Li, N., & Liu, J. (2010). Raman spectroscopic study for the determination of Cl⁻ concentration (molarity scale) in aqueous solutions: application to fluid inclusions. *Chemical Geology*, 272, 55-61.
- Tatirat, O., Charoenrein, S., & Kerr, W. L. (2012). Physicochemical properties of extrusion-modified konjac glucomannan. *Carbohydrate Polymers*, 87, 1545-1551.
- Wang, L. X., Jiang, Y. P., Lin, Y. H., Pang, J., & Liu, X. Y. (2016). Rheological properties and formation mechanism of DC electric fields induced konjac glucomannan-tungsten gels. *Carbohydrate Polymers*, 142, 293-299.
- Wang, W. J., Shen, M. Y., Liu, S. C., Jiang, L., Song, Q. Q., Xie, J. H. (2018). Gel properties and interactions of Mesona blumes polysaccharide-soy protein isolates mixed gel: The effect of salt addition. *Carbohydrate Polymers*, 192, 193-201.
- Wang, H. Y., Wan, L., Chen, D., Guo, X. F., Liu, F. X., & Pan, S. Y. (2019). Unexpected gelation behavior of citrus pectin induced by monovalent cations under alkaline conditions. *Carbohydrate Polymers*, 212, 51-58.
- Wang, K., Wu, K., Xiao, M., Kuang, Y., Corke, H., Ni, X. W., & Jiang, F. T. (2017). Structural characterization and properties of konjac glucomannan and zein blend films. *International Journal of Biological Macromolecules*, 105, 1096-1104.
- Wang, Y. X., Wu, K., Xiao, M., Riffat, S. B., Su, Y. H., & Jiang, F. T. (2018). Thermal conductivity, structure and mechanical properties of konjac glucomannan/starch based aerogel strengthened by wheat straw. *Carbohydrate Polymers*, 197, 284-291.
- Wang, X., Xiong, Y. L. L., & Sato, H. (2017). Rheological enhancement of pork myofibrillar protein-lipid emulsion composite gels via glucose oxidase oxidation/transglutaminase cross-linking pathway. *Journal of Agricultural and Food Chemistry*, 65 (38), 8451-8458.
- Wang, L. X., Zhuang, Y. H., Li, J. L., Pang, J., & Liu, X. Y. (2016). The textural properties and microstructure of konjac glucomannan-tungsten gels induced by DC electric fields. *Food Chemistry*, 212, 256-263.
- Wang, S. S., Zhou, B., Wang, Y. T., & Li, B. (2015). Preparation and characterization of konjac glucomannan microcrystals through acid hydrolysis. *Food Research International*, 67, 111-116.
- Xu, X. L., Han, M. Y., Fei, Y., & Zhou, G. H. (2011). Raman spectroscopic study of heat-induced gelation of pork myofibrillar proteins and its relationship with textural characteristic. *Meat Science*, 87(3), 159-164.
- Xue, S. W., Qian, C., Kim, H. Y. B., Xu, X. L., & Zhou, G. H. (2018). High-pressure effects on myosin in relation to heat gelation: A micro-perspective study, *Food Hydrocolloids*, 84, 219-228.
- Ye, S. X., Jin, W. P., Huang, Q., Hu, Y., Li, Y., Li, J., & Li, B. (2017). Da-KGM based GO-reinforced FMBO-loaded aerogels for efficient arsenic removal in aqueous solution. *International Journal of Biological Macromolecules*, 94, 527-534.
- Zhang, Y. Q., Xie, B. J., & Gan, X. (2005). Advance in the applications of konjac glucomannan and its derivatives, *Carbohydrate Polymers*, 60, 27-31.
- Zhang, M., Yang, X., Zhou, Z. Y., & Ye, X. Y. (2013). Controllable growth of gold nanowires and nanoactuators via high-frequency AC electrodeposition. *Electrochemistry Communications*, 27, 133-136.
- Zhang, K. G., Yang, W. Z., Xu, B., Chen, Y., Yin, X. S., Liu, Y., & Zuo, H. Z. (2018). Inhibitory effect of konjac

454 glucomanan on pitting corrosion of AA5052 aluminium alloy in NaCl solution, *Journal of Colloid and*
455 *Interface Science*, 517, 52-60.
456 Zhu, F. (2018). Modifications of konjac glucomannan for diverse applications. *Food Chemistry*, 256, 419-426.



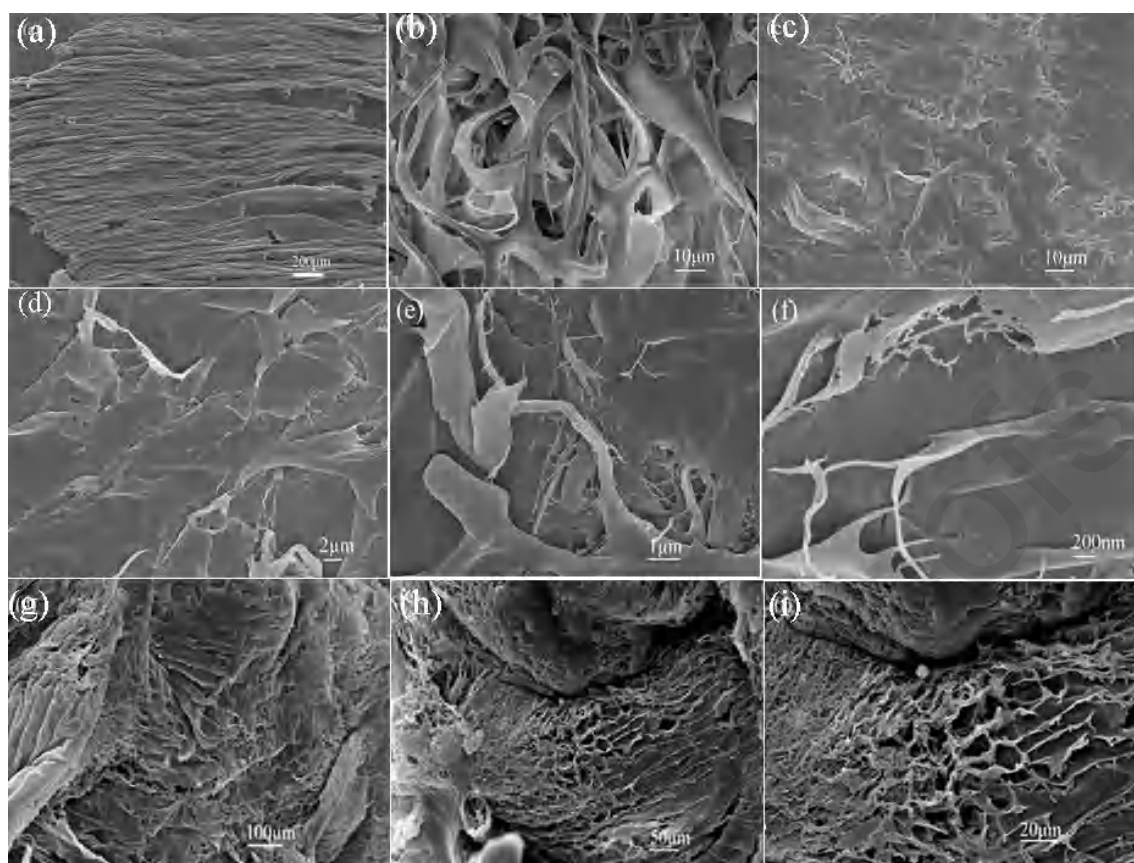
457
458

Fig. 1 FTIR spectra of native KGM, KGM-AC and KGM-KCl gel



459
460

Fig. 2 Raman spectra of KGM, KGM-AC and KGM-KCl gel

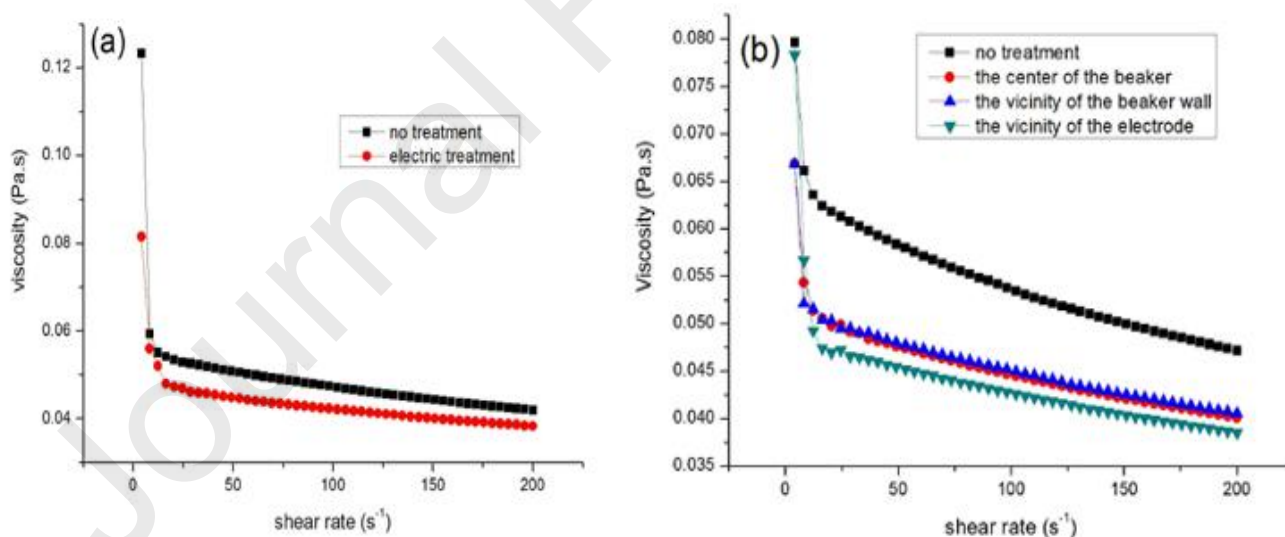


461

462

463

Fig. 3 SEM images of native KGM powder (a), lyophilized KGM hydrosol (b), KGM-AC (c-f) and lyophilized KGM-KCl gels (g-i) with different magnifications



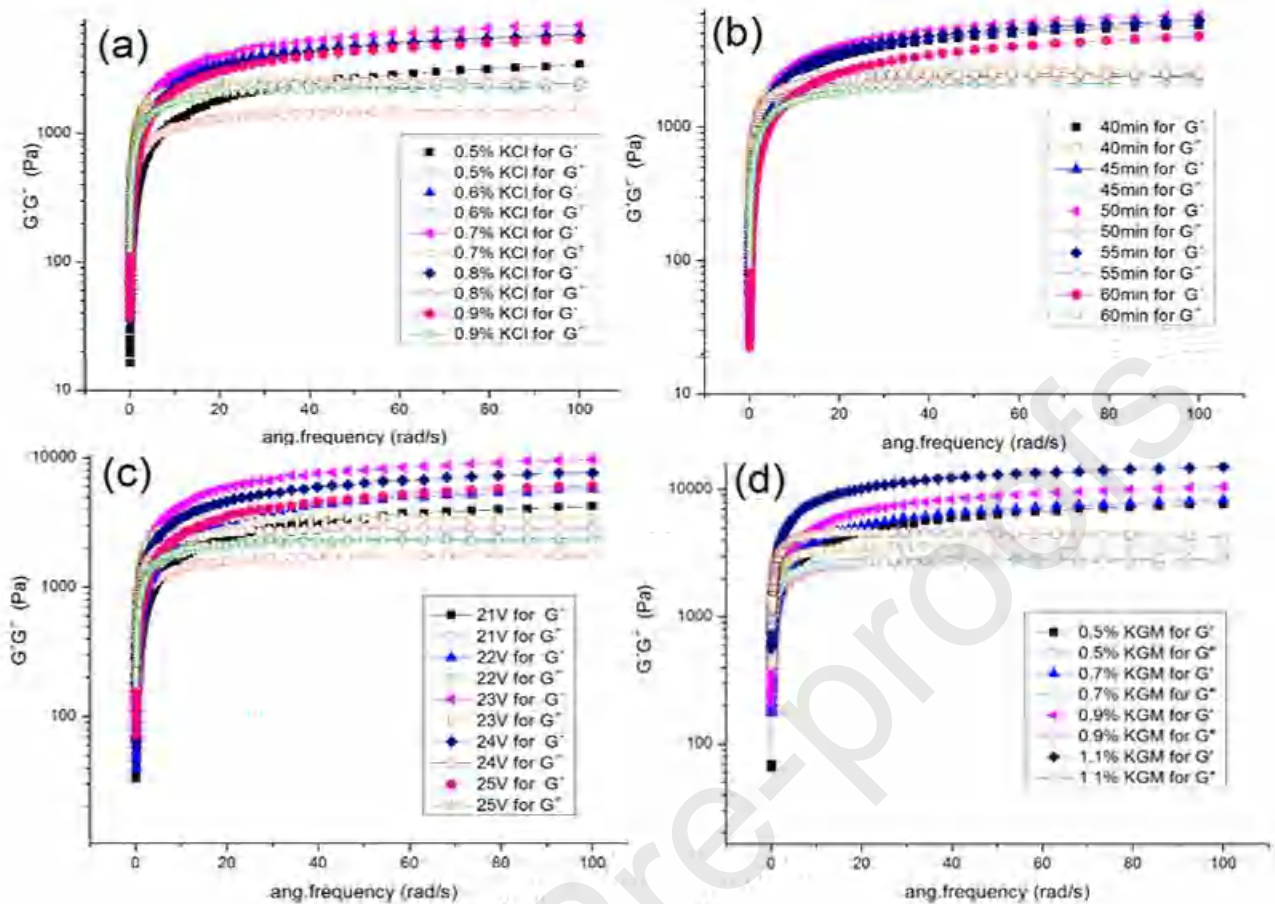
464

465

466

467

Fig. 4 Effect of an AC electric treatment on the apparent viscosity of KGM sols (a) and KGM-KCl mixed sols (b), a represents 0.5% KGM sol without and with electric treatment time for 30 min at 25 V, b represents 0.5% KGM sol containing 0.7% KCl without and with electric treatment time for 25 min at 25V



468

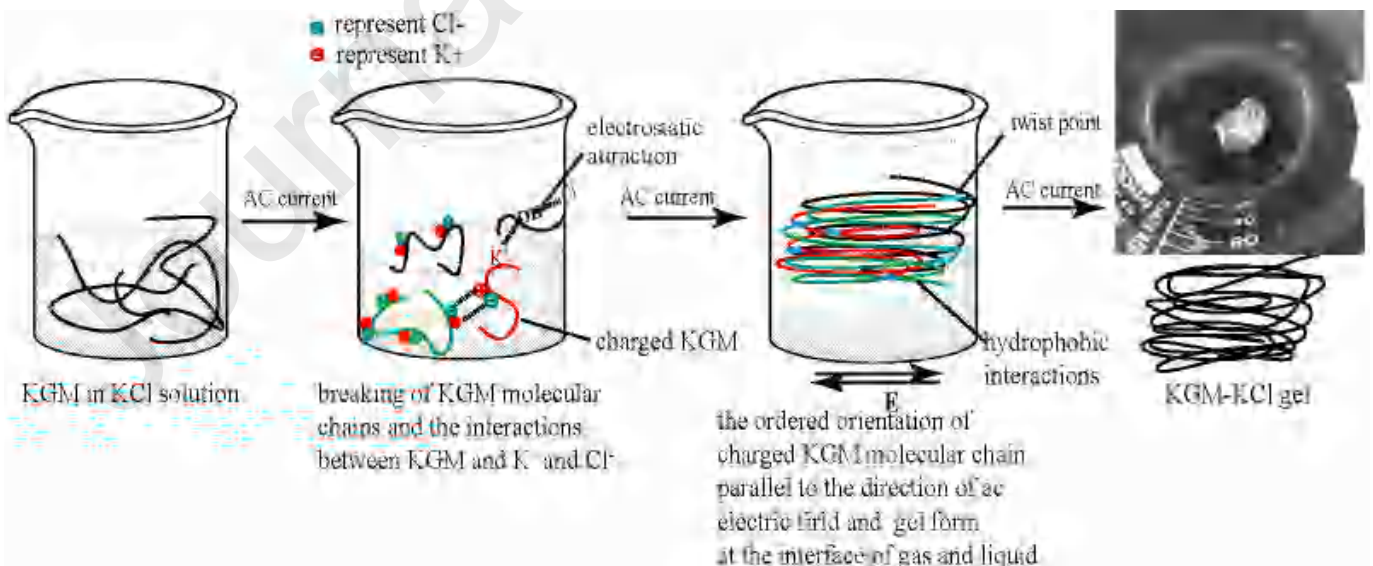
469

470

471

472

Fig.5 Frequency dependence of G' (full symbols) and G'' (open symbols) for KGM-KCl gels (a: different KCl concentration, c_{KGM} : 0.5%, voltage: 25V, time: 40min; b: different electric treatment time, c_{KGM} : 0.5%, c_K : 0.7%, voltage: 25V; c: different voltage, c_{KGM} : 0.5%, c_K : 0.7%, electric treatment time: 50min; d: different KGM concentration, c_K : 0.7%, electric treatment time: 50min, voltage: 23V)



473

474

475

476

Fig. 6 Schematic representation of the formation mechanism of KGM-KCl electrogel

477

478 **Ting-Jang Lu:** corresponding author, ensure that the descriptions are accurate and agreed by all authors.

479 **Li-Xia Wang:** Conceptualization, Methodology, Validation, Formal analysis, Investigation, Writing - Original

480 Draft, Writing - Review & Editing, Visualization.

481 **Anng-Ruei Lee:** Investigation.

482 **Yi Yuan:** Investigation.

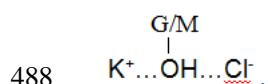
483 **Xiu-Mei Wang:** Investigation, Funding acquisition.

484

485

486

487 1.Potassium chloride interacts with hydroxyl groups of konjac glucomannan to form the structure of



489 2.Konjac glucomannan molecular chains break and partially deacetylate under alternating current electric field.

490 3.Alternating current induces the formation of konjac glucomannan-potassium chloride electrogel.

491 4.Konjac glucomannan-potassium chloride electrogel possesses an ununiform porosity structure.

492 5.Viscoelastic moduli of the gels depend on potassium chloride and konjac glucomannan concentration, voltage
493 and electric processing time.

494